

COX Inhibitor Screening Assay Kit

Item No. 560131



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GENERAL INFORMATION

Materials Supplied

Kit will arrive packaged as a -80°C kit. For best results, remove components and store as stated below.

Item Number	Item	Quantity	Storage
414016	PG Screening EIA Antiserum	1 vial	-20°C
414006	PG Screening AChE Tracer	1 vial	-20°C
414026	PG Screening EIA Standard	1 vial	-20°C
400060	EIA Buffer Concentrate (10X)	2 vials	Room temperature
400062	Wash Buffer Concentrate (400X)	1 vial	Room temperature
400035	Polysorbate 20	1 vial	Room temperature
400004	Mouse Anti-Rabbit IgG Coated Plate	1 plate	4°C
400012	96-Well Cover Sheet	1 cover	Room temperature
400050	Ellman's Reagent	3 vials	-20°C
460104	Reaction Buffer (10X)	1 vial	-20°C
460100	COX-1 (ovine)	1 vial	-80°C
460121	COX-2 (human recombinant)	1 vial	-80°C
460102	Heme	1 vial	-20°C
460103	Arachidonic Acid (substrate)	1 vial	-20°C
460105	Potassium Hydroxide	1 vial	-20°C
460106	Hydrochloric Acid	1 vial	-20°C
460107	Stannous Chloride	1 vial	-20°C

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.



WARNING: This product is for laboratory research use only: not for administration to humans. Not for human or veterinary diagnostic or therapeutic use.

Precautions

Please read these instructions carefully before beginning this assay.

The reagents in this kit have been tested and formulated to work exclusively with Cayman Chemical's ACE™ EIA Kits. This kit may not perform as described if any reagent or procedure is replaced or modified.

For research use only. Not for human or diagnostic use.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888
Fax: 734-971-3641
Email: techserv@caymanchem.com
Hours: M-F 8:00 AM to 5:30 PM EST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored as specified in the **Materials Supplied** section, on page 3, and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A plate reader capable of measuring absorbance between 405-420 nm.
2. Adjustable pipettes and a repeating or multichannel pipettor.
3. A source of 'UltraPure' water. Water used to prepare all EIA reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). Use activated carbon filter cartridges or other organic scavengers. Glass distilled water (even if double distilled), HPLC-grade water, and sterile water (for injections) are not adequate for EIA. *NOTE: UltraPure water is available for purchase from Cayman (Item No. 400000).*
4. A 37°C water bath or heat block.
5. Reaction tubes.
6. Materials used for purification procedure (optional, see page 30).

Background

Cyclooxygenase (COX, also called Prostaglandin H Synthase or PGHS) is a bifunctional enzyme exhibiting both COX and peroxidase activities. The COX component converts arachidonic acid to a hydroperoxy endoperoxide (PGG₂), and the peroxidase component reduces the endoperoxide to the corresponding alcohol (PGH₂), the precursor of prostaglandins (PGs), thromboxanes, and prostacyclins.^{1,2}

It is now well established that there are two distinct isoforms of COX. COX-1 is constitutively expressed in a variety of cell types and is involved in normal cellular homeostasis. A variety of stimuli, such as phorbol esters, lipopolysaccharides, and cytokines, lead to the induced expression of a second isoform of COX, COX-2. COX-2 is responsible for the biosynthesis of PGs under acute inflammatory conditions.^{3,4} This inducible COX-2 is believed to be the target enzyme for the anti-inflammatory activity of nonsteroidal anti-inflammatory drugs.⁴

About This Assay

The COX (ovine) Inhibitor Screening Assay directly measures PGF_{2α} by SnCl₂ reduction of COX-derived PGH₂ produced in the COX reaction. The prostanoid product is quantified *via* enzyme immunoassay (EIA) using a broadly specific antiserum that binds to all the major PG compounds.

This assay includes both ovine COX-1 and human recombinant COX-2 enzymes allowing the user to screen isozyme-specific inhibitors. This assay is an excellent tool which can be used for general inhibitor screening, or to eliminate false positive leads generated by less specific methods.

Description of ACE™ Competitive EIAs^{5,6}

This assay is based on the competition between PGs and a PG-acetylcholinesterase (AChE) conjugate (PG tracer) for a limited amount of PG antiserum. Because the concentration of the PG tracer is held constant while the concentration of PG varies, the amount of PG tracer that is able to bind to the PG antiserum will be inversely proportional to the concentration of PG in the well. This rabbit antiserum-PG (either free or tracer) complex binds to a mouse monoclonal anti-rabbit antibody that has been previously attached to the well. The plate is washed to remove any unbound reagents and then Ellman's Reagent (which contains the substrate to AChE) is added to the well. The product of this enzymatic reaction has a distinct yellow color and absorbs strongly at 412 nm. The intensity of this color, determined spectrophotometrically, is proportional to the amount of PG tracer bound to the well, which is inversely proportional to the amount of free PG present in the well during the incubation; or

$$\text{Absorbance} \propto [\text{Bound PG Tracer}] \propto 1/[\text{PG}]$$

A schematic of this process is shown in Figure 1, below.

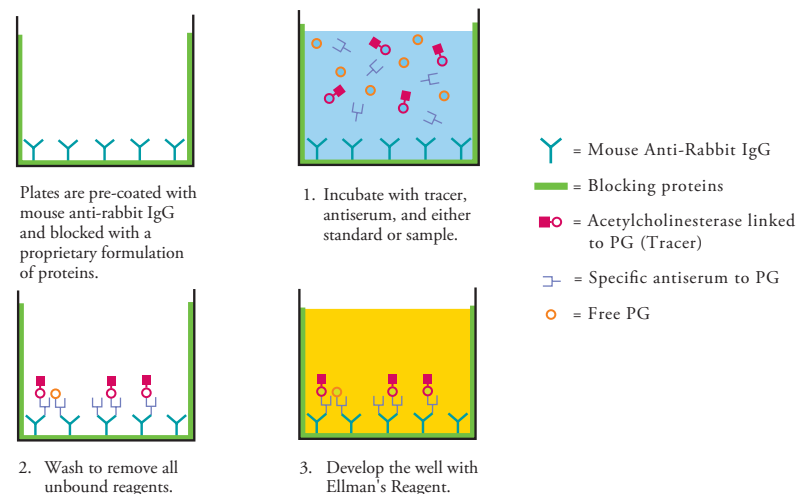


Figure 1. Schematic of the ACE™ EIA

Biochemistry of Acetylcholinesterase

The electric organ of the electric eel, *E. electricus*, contains an avid AChE capable of massive catalytic turnover during the generation of its electrochemical discharges. The electric eel AChE has a clover leaf-shaped tertiary structure consisting of a triad of tetramers attached to a collagen-like structural fibril. This stable enzyme is capable of high turnover ($64,000\text{ s}^{-1}$) for the hydrolysis of acetylthiocholine.

A molecule of the analyte covalently attached to a molecule of AChE serves as the tracer in ACE™ enzyme immunoassays. Quantification of the tracer is achieved by measuring its AChE activity with Ellman's Reagent. This reagent consists of acetylthiocholine and 5,5'-dithio-bis-(2-nitrobenzoic acid). Hydrolysis of acetylthiocholine by AChE produces thiocholine (see Figure 2, on page 9). The non-enzymatic reaction of thiocholine with 5,5'-dithio-bis-(2-nitrobenzoic acid) produces 5-thio-2-nitrobenzoic acid, which has a strong absorbance at 412 nm ($\epsilon = 13,600$).

AChE has several advantages over other enzymes commonly used for enzyme immunoassays. Unlike horseradish peroxidase, AChE does not self-inactivate during turnover. This property of AChE also allows redevelopment of the assay if it is accidentally splashed or spilled. In addition, the enzyme is highly stable under the assay conditions, has a wide pH range (pH 5-10), and is not inhibited by common buffer salts or preservatives. Since AChE is stable during the development step, it is unnecessary to use a 'stop' reagent, and the plate may be read whenever it is convenient.

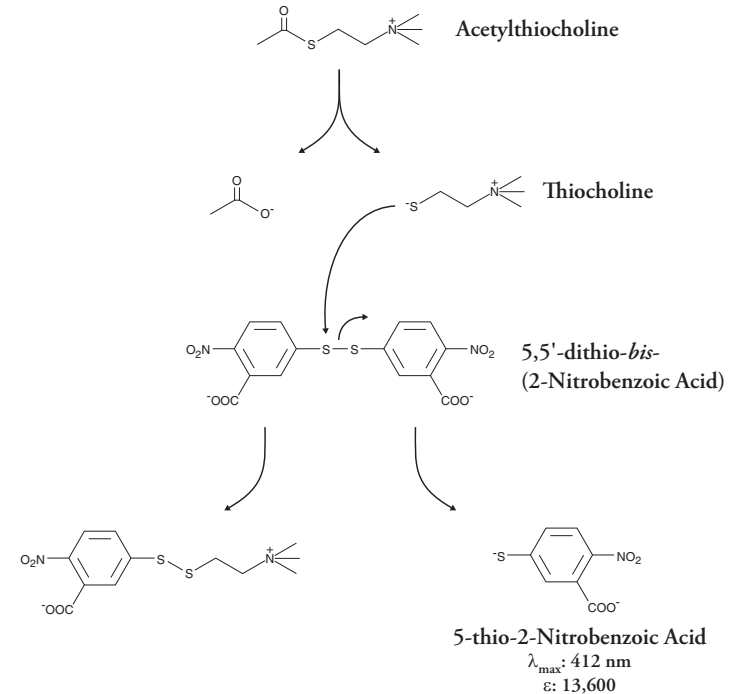


Figure 2. Reaction catalyzed by acetylcholinesterase

COX REACTION PROCEDURE

The use of both enzymes is not a requirement of the assay; one or both enzymes may be used depending on the nature of the study. The EIA plate will allow for 36 COX reactions (in duplicate) at one dilution or 18 COX reactions (in duplicate) at two dilutions.

IMPORTANT: Please read both COX Reaction Procedure and EIA Procedure sections carefully before initiating your experiments!

COX Reagent Preparation

1. Reaction Buffer (10X) - (Item No. 460104)

Dilute 5 ml of Reaction Buffer concentrate with 45 ml of UltraPure water. This final Reaction Buffer (0.1 M Tris-HCl, pH 8.0, containing 5 mM EDTA and 2 mM phenol) is used in the COX reactions and for dilution of Heme. When stored at room temperature, this diluted Reaction Buffer is stable for at least one month. Equilibrate the diluted Reaction Buffer to 37°C before using in the COX reactions.

2. COX-1 (ovine) - (Item No. 460100)

This vial contains a solution of ovine COX-1 and should be kept on ice when thawed. Dilute 80 µl of enzyme with 320 µl of diluted Reaction Buffer and store on ice. This quantity is sufficient for 40 reactions. The diluted enzyme is stable for one hour on ice. If less than 40 reactions will be performed, it is recommended that the undiluted enzyme be aliquoted into several small vials and stored at -80°C to avoid repeated freeze thaw cycles.

3. COX-2 (human recombinant) - (Item No. 460121)

This vial contains a solution of human recombinant COX-2 and should be kept on ice when thawed. Dilute 80 µl of enzyme with 320 µl of diluted Reaction Buffer and store on ice. This quantity is sufficient for 40 reactions. The diluted enzyme is stable for one hour on ice. If less than 40 reactions will be performed, it is recommended that the undiluted enzyme be aliquoted into several small vials and stored at -80°C to avoid repeated freeze thaw cycles.

4. Heme - (Item No. 460102)

This vial contains a solution of Heme in dimethylsulfoxide (DMSO). Dilute 40 µl of Heme with 960 µl of 1X Reaction Buffer prior to use. The diluted Heme is stable for 12 hours at room temperature.

5. Arachidonic Acid (Substrate) - (Item No. 460103)

This vial contains a solution of Arachidonic Acid in ethanol. Transfer 50 µl of the supplied Substrate to another vial, add 50 µl of Potassium Hydroxide (Item No. 460105), vortex, and dilute with 400 µl of UltraPure water to achieve a final concentration of 2 mM. Use the prepared Arachidonic Acid Solution within one hour. A 10 µl aliquot of the prepared substrate will yield a final concentration of 100 µM in the reaction.

6. Potassium Hydroxide - (Item No. 460105)

This vial contains 0.1 M Potassium Hydroxide (KOH). The reagent is ready to use as supplied.

7. Hydrochloric Acid - (Item No. 460106)

This vial contains 1 M Hydrochloric Acid (HCl). The HCl is used to prepare the saturated Stannous Chloride Solution.

8. Stannous Chloride - (Item No. 460107)

This vial contains crystalline Stannous Chloride. Add 5 ml of Hydrochloric Acid (Item No. 460106) and vortex to produce a saturated solution of Stannous Chloride. (This saturated Stannous Chloride Solution may be cloudy.) This solution is stable for eight hours at room temperature. If not performing all of the COX reactions in one day, weigh 125 mg of Stannous Chloride into another vial and add 2.5 ml of Hydrochloric Acid (Item No. 460106). 30 µl of saturated Stannous Chloride is required for each reaction. *NOTE: Stannous Chloride is used to reduce PGH₂, produced in the COX reaction, to a more stable PG, PGF_{2α}*

Performing COX Reactions

Pipetting Hints

- It is recommended that a repeating or multichannel pipettor be used to deliver Arachidonic Acid to the reaction tubes. This saves time and helps to maintain more precise incubation times.
 - Use different tips to pipette the buffer, enzyme, Heme, inhibitor, and Arachidonic Acid.
 - Before pipetting each reagent, equilibrate the pipette tip in that reagent (*i.e.*, slowly fill the tip and gently expel the contents, repeat several times).
 - Do not expose the pipette tip to the reagent(s) already in the reaction tube.
-
- *The final volume of the reaction is 230 µl in all the reaction tubes.*
 - *You do not have to use both enzymes. You can use either COX-1 or COX-2 depending on your experimental design. The EIA plate will allow for 36 COX reactions (in duplicate) at one dilution or 18 COX reactions (in duplicate) at two dilutions.*
 - *Use the 1X Reaction Buffer in the reactions and pre-equilibrate to 37°C.*
 - *Set the water bath or heat block temperature to 37°C before initiating the reactions.*
 - *It is recommended that no more than 18 reactions be performed at one time.*
 - *Aliquot reagents into reaction tubes that are at 37°C.*
1. **Background tubes** - Inactivate COX-1 and COX-2 by transferring 20 µl of each enzyme diluted to a 500 µl microfuge tube and placing the tube in boiling water for three minutes. The inactivated enzymes will be used to generate the background values. Add the following reagents to two reaction tubes: 160 µl of Reaction Buffer, 10 µl of Heme, and 10 µl of inactive COX-1 or inactive COX-2.
 2. **COX-1 100% Initial Activity tubes** - Add 160 µl of Reaction Buffer, 10 µl of Heme, and 10 µl of COX-1 to two reaction tubes.
 3. **COX-1 Inhibitor tubes** - Add 160 µl of Reaction Buffer, 10 µl of Heme, and 10 µl of COX-1 to six reaction tubes.
 4. **COX-2 100% Initial Activity tubes** - add 160 µl of Reaction Buffer, 10 µl of Heme, and 10 µl of COX-2 to two reaction tubes.

5. **COX-2 Inhibitor tubes** - add 160 µl of Reaction Buffer, 10 µl of Heme, and 10 µl of COX-2 to six reaction tubes.
6. Add 10 µl of inhibitor* to the COX-1 and -2 inhibitor tubes and 10 µl of inhibitor vehicle to the 100% Initial Activity and Background tubes.
7. Incubate for 10 minutes at 37°C. *NOTE: Most inhibitors exhibit time-dependent inhibition of COX activity. Altering incubation times with the inhibitor can significantly change the apparent IC₅₀ value of the compound. Determining the optimal pre-incubation times for inhibitors is suggested.*
8. Initiate the reaction by adding 10 µl of Arachidonic Acid to all the reaction tubes. Quickly mix and incubate for EXACTLY two minutes at 37°C.
9. Add 30 µl of the saturated Stannous Chloride solution to each reaction tube to stop enzyme catalysis. Remove tubes from the water bath and vortex. Incubate for five minutes at room temperature. The reaction mixture will be cloudy.
10. Repeat steps 2-8 if performing more reactions.
11. The prostaglandins are quantified by EIA. Proceed to the **EIA Procedure**. The reactions are stable for one week at 0-4°C if tightly capped.

*Inhibitors can be dissolved in methanol, dimethylsulfoxide, or ethanol and should be added to the assay in a final volume of 10 µl. In the event that the appropriate concentration of inhibitor is completely unknown, we recommend that several dilutions of the inhibitor be made.

EIA PROCEDURE

NOTE: Water used to prepare all EIA reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). Use activated carbon filter cartridges or other organic scavengers. Glass distilled water (even if double distilled), HPLC-grade water, and sterile water (for injections) are not adequate for EIA. UltraPure water may be purchased from Cayman (Item No. 400000).

EIA Buffer Preparation

Store all diluted buffers at 4°C; they will be stable for about two months.

1. EIA Buffer Preparation

Dilute the contents of one vial of EIA Buffer Concentrate (10X) (Item No. 400060) with 90 ml of UltraPure water. Be certain to rinse the vial to remove any salts that may have precipitated. *NOTE: It is normal for the concentrated buffer to contain crystalline salts after thawing. These will completely dissolve upon dilution with water.*

2. Wash Buffer Preparation

Dilute the contents of the vial of Wash Buffer Concentrate (400X) (Item No. 400062) to a total volume of 2 liters with UltraPure water and add 1 ml of Polysorbate 20 (Item No. 400035). A smaller volume of Wash Buffer can be prepared by diluting the Wash Buffer Concentrate 1:400 and adding Polysorbate 20 (0.5 ml/L of Wash Buffer). The diluted buffer will be stable for two months at 4°C.

NOTE: Polysorbate 20 is a viscous liquid and cannot be measured by a pipette. A positive displacement device such as a syringe should be used to deliver small quantities accurately.

Preparation of Assay-Specific Reagents

Prostaglandin Screening Standard

Dissolve the lyophilized PG Screening EIA Standard (Item No. 414026) in 1 ml of EIA Buffer. The concentration of this solution (the bulk standard) will be 10 ng/ml. Store this solution at 4°C; it will be stable for approximately six weeks.

To prepare the standard for use in EIA: Obtain eight clean test tubes and number them #1 through #8. Aliquot 800 µl EIA Buffer to tube #1 and 500 µl EIA Buffer to tubes #2-8. Transfer 200 µl of the bulk standard (10 ng/ml) to tube #1 and mix thoroughly. Serially dilute the standard by removing 500 µl from tube #1 and placing in tube #2; mix thoroughly. Next, remove 500 µl from tube #2 and place into tube #3; mix thoroughly. Repeat this process for tubes #4-8. These diluted standards should not be stored for more than 24 hours.

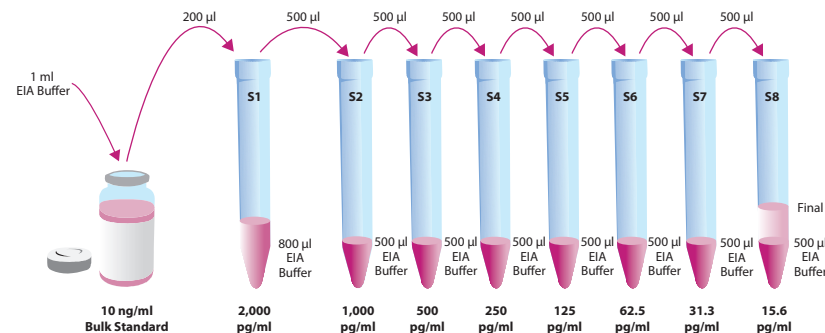


Figure 3. Preparation of the PG standards

Prostaglandin Screening AChE Tracer

Reconstitute the 100 dtn PG Screening AChE Tracer (Item No. 414006) with 6 ml EIA Buffer. Store the reconstituted PG Screening AChE Tracer at 4°C (*do not freeze!*) and use within two weeks. A 20% surplus of tracer has been included to account for any incidental losses.

Prostaglandin Screening EIA Antiserum

Reconstitute the 100 dtn PG Screening EIA Antiserum (Item No. 414016) with 6 ml EIA Buffer. Store the reconstituted PG Screening EIA Antiserum at 4°C. It will be stable for at least four weeks. A 20% surplus of antiserum has been included to account for any incidental losses.

COX Reaction Dilutions

Background Samples

Obtain two clean test tubes and label them BC1 and BC2. Aliquot 990 µl of EIA Buffer to each test tube. Add 10 µl of background COX-1 to the tube labeled BC1, 10 µl of background COX-2 to the tube labeled BC2, and mix thoroughly. Each test tube contains a 1:100 dilution of the original sample.

COX 100% Initial Activity Samples

Obtain three clean test tubes per sample and number them IA1 through IA3. Aliquot 990 µl of EIA Buffer to tube IA1, add 10 µl of COX-1 or COX-2 100% Initial Activity sample, and mix thoroughly. Aliquot 950 µl of EIA Buffer to tube IA2, add 50 µl of tube IA1 to tube IA2, and mix thoroughly. Tube IA2 contains a 1:2,000 dilution of the original sample. Aliquot 500 µl of EIA Buffer to tube IA3, add 500 µl of tube IA2 to tube IA3, and mix thoroughly. Tube IA3 contains a 1:4,000 dilution of the original sample. Tubes IA2 and IA3 will be run in the EIA. Do not use test tube IA1 in the EIA, this tube is too concentrated (this dilution is outside the usable range of the assay).

COX Inhibitor Samples

Obtain three clean test tubes per sample and number them C1 through C3. Aliquot 990 µl of EIA Buffer to tube C1, add 10 µl of sample, and mix thoroughly. Aliquot 950 µl of EIA Buffer to tube C2, add 50 µl of tube C1 to tube C2, and mix thoroughly. Tube C2 contains a 1:2,000 dilution of the original sample. Aliquot 500 µl of EIA Buffer to tube C3, add 500 µl of tube C2 to tube C3, and mix thoroughly. Tube C3 contains a 1:4,000 dilution of the original sample. Tubes C2 and C3 will be run in the EIA. Do not use test tube C1 in the EIA, this tube is too concentrated (this dilution is outside the usable range of the assay).

Definition of Key Terms

Blank: background absorbance caused by Ellman's Reagent. The blank absorbance should be subtracted from the absorbance readings of all the other wells, including NSB wells.

Total Activity: total enzymatic activity of the AChE-linked tracer. This is analogous to the specific activity of a radioactive tracer.

NSB (Non-Specific Binding): non-immunological binding of the tracer to the well. Even in the absence of specific antibody a very small amount of tracer still binds to the well; the NSB is a measure of this low binding. Do not forget to subtract the Blank absorbance values.

B₀ (Maximum Binding): maximum amount of the tracer that the antibody can bind in the absence of free analyte.

%B/B₀ (%Bound/Maximum Bound): ratio of the absorbance of a particular sample or standard well to that of the maximum binding (B₀) well.

Standard Curve: a plot of the %B/B₀ values *versus* concentration of a series of wells containing various known amounts of analyte.

Dtn: determination, where one dtn is the amount of reagent used per well.

Plate Set Up

The 96-well plate(s) included with this kit is supplied ready to use. It is not necessary to rinse the plate(s) prior to adding the reagents. *NOTE: If you do not need to use all the strips at once, place the unused strips back in the plate packet and store at 4°C. Be sure the packet is sealed with the desiccant inside.*

Each plate or set of strips must contain a minimum of two Blanks (Blk), two Non-Specific Binding wells (NSB), two Maximum Binding wells (B_0), one Total Activity well (TA), and an eight point standard curve run in duplicate. *NOTE: Each assay must contain this minimum configuration in order to ensure accurate and reproducible results.* Each COX reaction sample should be assayed at two dilutions and each dilution should be assayed in duplicate. A minimum of one 100% Initial Activity Sample should be assayed for both COX-1 and -2.

A suggested plate format is shown in Figure 4, below. The user may vary the location and type of wells present as necessary for each particular experiment. The plate format provided below has been designed to allow for easy data analysis using a convenient spreadsheet offered by Cayman (see page 22, for more details). We suggest you record the contents of each well on the template sheet provided (see page 35).

Blk - Blank
TA - Total Activity
NSB - Non-Specific Binding
 B_0 - Maximum Binding
S1-S8 - Standards 1-8
BC1 - Background COX-1
BC2 - Background COX-2
‡ - 100% Initial Activity Samples (IA2 & IA3)
H - COX Inhibitor Samples (C2 & C3)

	1	2	3	4	5	6	7	8	9	10	11	12
A	Blk	S1	S1	BC1	BC1	H	H	H	H	H	H	H
B	Blk	S2	S2	BC2	BC2	H	H	H	H	H	H	H
C	NSB	S3	S3	‡	‡	H	H	H	H	H	H	H
D	NSB	S4	S4	‡	‡	H	H	H	H	H	H	H
E	B_0	S5	S5	‡	‡	H	H	H	H	H	H	H
F	B_0	S6	S6	‡	‡	H	H	H	H	H	H	H
G	B_0	S7	S7	H	H	H	H	H	H	H	H	H
H	TA	S8	S8	H	H	H	H	H	H	H	H	H

Figure 4. Sample plate format

Performing the EIA

Pipetting Hints

- Use different tips to pipette each reagent.
- Before pipetting each reagent, equilibrate the pipette tip in that reagent (*i.e.*, slowly fill the tip and gently expel the contents, repeat several times).
- Do not expose the pipette tip to the reagent(s) already in the well.

Addition of the Reagents

1. EIA Buffer

Add 100 μ l of EIA Buffer to NSB wells. Add 50 μ l of EIA Buffer to B_0 wells.

2. Prostaglandin Screening EIA Standard

Add 50 μ l from tube #8 to both of the lowest standard wells (S8). Add 50 μ l from tube #7 to each of the next two standard wells (S7). Continue with this procedure until all the standards are aliquoted. The same pipette tip should be used to aliquot all the standards. Before pipetting each standard, be sure to equilibrate the pipette tip in that standard.

3. Background Samples (test tubes BC1 and BC2)

Add 50 μ l of sample per well. Each sample should be assayed in duplicate.

4. COX 100% Initial Activity Samples (test tubes IA2 and IA3)

Add 50 μ l of sample per well. We recommend that you assay each sample at the two dilutions with each dilution assayed in duplicate.

5. COX Inhibitor Samples (test tubes C2 and C3)

Add 50 μ l of sample per well. We recommend that you assay each sample at the two dilutions with each dilution assayed in duplicate.

6. Prostaglandin Screening AChE Tracer

Add 50 μ l of PG Screening AChE Tracer to each well *except* the TA and the Blk wells.

7. Prostaglandin Screening EIA Antiserum

Add 50 µl of PG Screening EIA Antiserum to each well *except* the TA, the NSB, and the Blk wells.

Well	EIA Buffer	Standard/ Sample	Tracer	Antiserum
Blk	-	-	-	-
TA	-	-	5 µl (at devl. step)	-
NSB	100 µl	-	50 µl	-
B ₀	50 µl	-	50 µl	50 µl
Std/Sample	-	50 µl	50 µl	50 µl

Table 1. Pipetting summary

Incubation of the Plate

Cover each plate with plastic film (Item No. 400012) and incubate for 18 hours at room temperature on an orbital shaker.

Development of the Plate

- When ready to develop the plate, reconstitute one 100 dtn vial of Ellman’s Reagent (Item No. 400050) with 20 ml of UltraPure water. This reagent is sufficient to develop 100 wells.

NOTE: Reconstituted Ellman’s Reagent is unstable and should be used the same day it is prepared; protect the Ellman’s Reagent from light when not in use. Extra vials of the reagent have been provided should a plate need to be re-developed or multiple assays run on different days.

- Empty the wells and rinse five times with Wash Buffer.
- Add 200 µl of Ellman’s Reagent to each well.
- Add 5 µl of tracer to the TA wells.
- Cover the plate with plastic film. Optimum development is obtained by using an orbital shaker equipped with a large, flat cover to allow the plate to develop in the dark. This assay typically develops (*i.e.*, B₀ wells ≥0.3 A.U. (blank subtracted)) in 60-90 minutes.

Reading the Plate

- Wipe the bottom of the plate with a clean tissue to remove fingerprints, dirt, etc.
- Remove the plate cover being careful to keep Ellman’s Reagent from splashing on the cover. *NOTE: Any loss of Ellman’s Reagent will affect the absorbance readings. If Ellman’s Reagent is present on the cover, use a pipette to transfer the Ellman’s Reagent into the well. If too much Ellman’s Reagent has splashed on the cover to easily redistribute back into the wells, wash the plate three times with wash buffer and repeat the development with fresh Ellman’s Reagent.*
- Read the plate at a wavelength between 405 and 420 nm. The absorbance may be checked periodically until the B₀ wells have reached a minimum of 0.3 A.U. (blank subtracted). The plate should be read when the absorbance of the B₀ wells are in the range of 0.3-0.8 A.U. (blank subtracted). If the absorbance of the wells exceeds 1.5, wash the plate, add fresh Ellman’s Reagent and let it develop again.

ANALYSIS

Many plate readers come with data reduction software that plot data automatically. Alternatively a spreadsheet program can be used. The data should be plotted as %B/B₀ versus log concentration using either a 4-parameter logistic or log-logit curve fit. *NOTE: Cayman has a computer spreadsheet available for data analysis. Please contact Technical Service or visit our website (www.caymanchem.com/analysis/eia) to obtain a free copy of this convenient data analysis tool.*

Calculations

Preparation of the Data

The following procedure is recommended for preparation of the data prior to graphical analysis.

NOTE: If the plate reader has not subtracted the absorbance readings of the blank wells from the absorbance readings of the rest of the plate, be sure to do that now.

1. Average the absorbance readings from the NSB wells.
2. Average the absorbance readings from the B₀ wells.
3. Subtract the NSB average from the B₀ average. This is the corrected B₀ or corrected maximum binding.
4. Calculate the %B/B₀ (% Sample or Standard Bound/Maximum Bound) for the remaining wells. To do this, subtract the average NSB absorbance from the S1 absorbance and divide by the corrected B₀ (from Step 3). Multiply by 100 to obtain %B/B₀. Repeat for S2-S8 and all sample wells.

NOTE: The TA values are not used in the standard curve calculations. Rather, they are used as a diagnostic tool; the corrected B₀ divided by the actual TA (10X measured absorbance) will give the % Bound. This value should closely approximate the % Bound that can be calculated from the Sample Data (see page 25). Erratic absorbance values and a low (or no) % Bound could indicate the presence of organic solvents in the buffer or other technical problems (see page 31 for Troubleshooting).

If you have purified your samples (see **Interference**, page 29), the final sample concentrations can be determined as follows:

Calculations

$$\text{Recovery Factor} = \frac{10 \times \text{cpm of sample}}{[\text{H}]-\text{PGE}_2 \text{ added to sample (cpm)}}$$

$$\text{PG (pg) in purified sample} = \left[\frac{\text{Value from EIA (pg/ml)}}{\text{Recovery Factor}} \right] \times 0.5 \text{ ml} - \text{added } [\text{H}]-\text{PGE}_2 \text{ (pg)}$$

$$\text{Total PG in sample (pg/ml)} = \frac{\text{PG (pg) in purified sample}}{\text{Volume of sample used for purification (ml)}}$$

Plot the Standard Curve

Plot %B/B₀ for standards S1-S8 versus Prostaglandin concentration using linear (y) and log (x) axes and perform a 4-parameter logistic fit.

Alternative Plot - The data can also be linearized using a logit transformation. The equation for this conversion is shown below. *NOTE: Do not use %B/B₀ in this calculation.*

$$\text{logit (B/B}_0\text{)} = \ln [\text{B/B}_0\text{}/(1 - \text{B/B}_0\text{)}]$$

Plot the data as logit (B/B₀) versus log concentrations and perform a linear regression fit.

Determine the Sample Concentration

1. Calculate the %B/B₀ value for each sample.
2. Determine the concentration of each sample by identifying the %B/B₀ on the standard curve and reading the corresponding values on the x-axis. %B/B₀ values greater than 80% and less than 20% should be re-assayed as they generally fall out of the linear range of the standard curve. A 20% or greater disparity between the apparent concentration of two different dilutions of the same sample could indicate interference which may be eliminated by purification. Remember to multiply the COX samples by the appropriate dilution factor (BC1 and BC2 = 100; IA2 and C2 = 2,000; IA3 and C3 = 4,000).
3. Subtract the background values (BC1 and BC2) from the 100% Initial Activity and COX Inhibitor samples.
4. Subtract each Inhibitor Sample from the 100% Initial Activity Sample, then divide by the 100% Initial Activity Sample, and multiply by 100 to give the percent inhibition.
5. Graph the percent inhibition by the inhibitor concentration to determine the IC₅₀ value (concentration at which there was 50% inhibition).

EIA Performance Characteristics

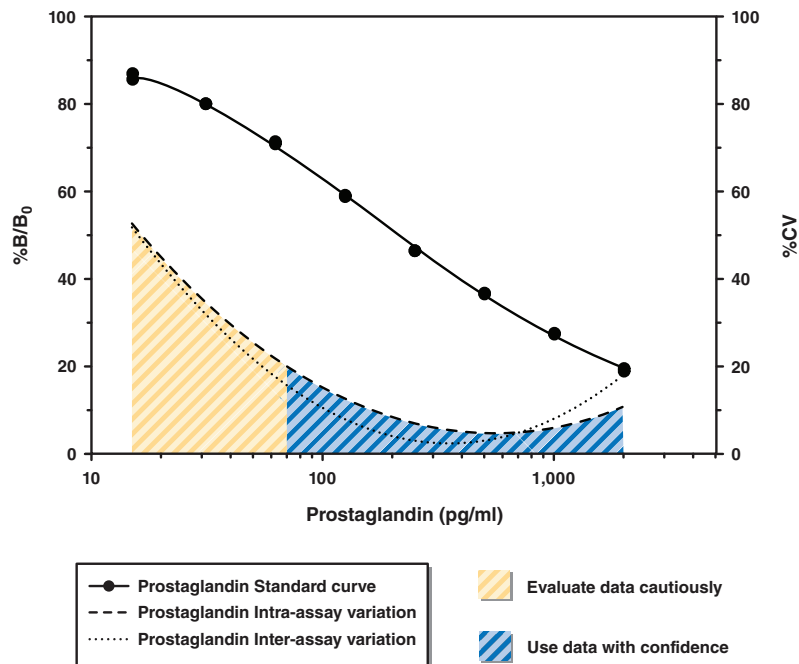
Sample Data

The standard curve presented here is an example of the data typically produced with this kit; however, your results will not be identical to these. You must run a new standard curve. Do not use the data below to determine the value of your samples. Your results could differ substantially.

	Raw Data		Average	Corrected		
Total Activity	1.850	2.019	1.935			
NSB	0.003	0.002	0.003			
B ₀	0.779	0.766				
	0.771	0.778	0.774	0.771		

Dose (pg/ml)	Raw Data		Corrected		%B/B ₀	
2,000	0.148	0.153	0.145	0.150	18.9	19.5
1,000	0.214	0.215	0.211	0.212	27.4	27.6
500	0.286	0.285	0.283	0.282	36.8	36.6
250	0.362	0.361	0.359	0.358	46.6	46.5
125	0.457	0.459	0.454	0.456	58.9	59.2
62.5	0.554	0.550	0.551	0.547	71.5	71.0
31.3	0.620	0.621	0.617	0.618	80.1	80.2
15.6	0.664	0.674	0.661	0.671	85.8	87.1

Table 2. Typical results



Precision:

The intra- and inter-assay CVs have been determined at multiple points on the standard curve. These data are summarized in the graph on page 26 and in the table below.

Dose (pg/ml)	%CV* Intra-assay variation	%CV* Inter-assay variation
2,000	7.9	13.1
1,000	8.1	9.8
500	8.2	6.8
250	8.6	9.7
125	10.6	10.2
62.5	11.0	8.8
31.3	†	19.4
15.6	†	†

Table 3. Intra- and inter-assay variation

*%CV represents the variation in concentration (not absorbance) as determined using a reference standard curve.

†Outside of the recommended usable range of the assay.

Specificity:

Compound	Cross Reactivity	Compound	Cross Reactivity
Prostaglandin E ₁	100%	Leukotriene D ₄	0.2%
Prostaglandin E ₂	100%	Arachidonic Acid	<0.01%
Prostaglandin F _{1α}	100%	Leukotriene B ₄	<0.01%
Prostaglandin F _{2α}	100%	Leukotriene C ₄	<0.01%
Prostaglandin F _{3α}	51.3%	Leukotriene E ₄	<0.01%
Prostaglandin E ₂ Ethanolamide	44.0%	Misoprostol	<0.01%
6-keto Prostaglandin F _{1α}	43.6%	Misoprostol (free acid)	<0.01%
8- <i>iso</i> Prostaglandin F _{2α}	38.4%	Prostaglandin A ₁	<0.01%
8- <i>iso</i> Prostaglandin E ₂	28.5%	Prostaglandin A ₂	<0.01%
Prostaglandin D ₂	26.6%	Prostaglandin A ₃	<0.01%
8- <i>iso</i> -2,3-dinor Prostaglandin F _{1α}	20.0%	Prostaglandin B ₁	<0.01%
Prostaglandin E ₃	9.5%	15-keto Prostaglandin E ₂	<0.01%
Thromboxane B ₂	5.0%	13,14-dihydro-15-keto Prostaglandin F _{2α}	<0.01%
12(S)-HHTrE	0.25%		

Table 4. Specificity of the PG Screening Antiserum

RESOURCES

Interference

It is possible that a COX inhibitor will interfere with the EIA and thus appear to exhibit no enzyme inhibition or exhibit a higher prostaglandin value than the 100% initial activity well. If the inhibitor exhibits no inhibition, you can either repeat the COX reaction using a higher concentration of inhibitor or purify the sample and repeat the EIA. If the sample exhibits a higher PG value than the 100% initial activity well, purify the sample and repeat the EIA. You can also test for inhibitor interference by adding the inhibitor to a boiled enzyme sample as a control. Treat the control as a normal COX Inhibitor Sample. The sample should not yield any PGs. If the inhibitor is detected by the antiserum, the inhibitor is interfering with the EIA.

To standardize the assay with specific COX inhibitors, use SC-560 (Item No. 70340) and DuP697 (Item No. 70645) for COX-1 and COX-2, respectively. At a final concentration of 20 nM SC-560 and 5 nM DuP-697 in the COX reactions, these inhibitors will yield approximately 50% inhibition.

Sample purification procedure to be followed only if an inhibitor is interfering with the assay:

1. Add 10,000 cpm of radiolabeled PGE₂ ([³H]-PGE₂) to the sample. We recommend that a high specific activity tracer be used in order to minimize the amount of radioactive PGE₂ added. The EIA will be able to detect the added PGE₂ and therefore the amount added should be insignificant in comparison to the endogenous analyte, yet should be sufficient for accurate scintillation counting.
2. Adjust the pH of the sample to ~4.0 using 1.0 M acetate buffer or citrate buffer (pH 4.0). (Standardize the pH adjustment using the sample matrix prior to proceeding with a large number of samples; approximately 1-2 equivalents of buffer is required for most biological samples.) If the samples are cloudy or contain precipitate, either filter or centrifuge to remove the precipitate. Particulate matter in the sample may clog the SPE (solid phase extraction) cartridge, resulting in loss of the sample.
3. Activate a SPE Cartridge (C-18) (6 ml) (Item No. 400020) by rinsing with 5 ml methanol and then with 5 ml UltraPure water. Do not allow the SPE cartridge to dry. *NOTE: Use one cartridge per sample.*
4. Pass the sample through the SPE cartridge. Rinse the cartridge with 5 ml UltraPure water. Discard the washes. Elute the PGE₂ with 5 ml ethyl acetate containing 1% methanol. Higher recovery and better reproducibility may be obtained if the sample is applied and eluted by gravity. The wash steps may be performed under vacuum or pressure.
5. Remove 500 µl for scintillation counting.
6. Evaporate the ethyl acetate/methanol to dryness either by vacuum centrifugation or by evaporation under a stream of dry nitrogen. It is imperative that all of the organic solvent is removed, as even trace quantities will adversely affect the EIA.
7. Add 1.035 ml of EIA Buffer and vortex. Proceed to **COX Reaction Dilutions** (see page 16) for the sample dilutions.

Troubleshooting

Problem	Possible Causes	Recommended Solutions
Erratic values; dispersion of duplicates	A. Trace organic contaminants in the water source B. Poor pipetting/technique	Replace activated carbon filter or change source of UltraPure water
High NSB (>0.035)	A. Poor washing B. Exposure of NSB wells to specific antibody	Rewash plate and redevelop
Very low B ₀	A. Trace organic contaminants in the water source B. Plate requires additional development time C. Dilution error in preparing reagents	A. Replace activated carbon filter or change source of UltraPure water B. Return plate to shaker and re-read later
Low sensitivity (shift in dose response curve)	Standard is degraded	Replace standard
Analyses of two dilutions do not agree (i.e., more than 20% difference)	Interfering substances are present	See Interference section on page 29
Only Total Activity (TA) wells develop	Trace organic contaminants in the water source	Replace activated carbon filter or change source of UltraPure water
No inhibition seen with compound	A. The concentration of the compound is not high enough B. The compound is not an inhibitor of the enzyme	Increase the compound concentration and re-assay

Additional Reading

Go to www.caymanchem.com/560131/references for a list of publications citing the use of Cayman's COX Inhibitor Screening Assay Kit.

References

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3. Xie, W., Chipman, J.G., Robertson, D.L., *et al.* Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing. *Proc. Natl. Acad. Sci. USA* **88**, 2692-2696 (1991).
4. Blobaum, A.L. and Marnett, L.J. Structural and functional basis of cyclooxygenase inhibition. *J. Med. Chem.* **50**(7), 1425-1441 (2007).
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Related Products

Arachidonic Acid - Item No. 90010
COX Activity Assay - Item No. 760151
COX Colorimetric Inhibitor Screening Assay Kit - Item No. 701050
COX Fluorescent Activity Assay Kit - Item No. 700200
COX Fluorescent Inhibitor Screening Assay Kit - Item No. 700100
COX (ovine) Colorimetric Inhibitor Screening Assay Kit - Item No. 760111
COX (ovine) Inhibitor Screening Assay Kit - Item No. 560101
COX-1 (ovine) - Item No. 60100
COX-1 (ovine) Inhibitor Screening Assay Kit - Item No. 701090
COX-2 (human) Inhibitor Screening Assay Kit - Item No. 701080
COX-2 (human recombinant) - Item No. 60122
COX-2 (ovine) - Item No. 60120
DuP-697 - Item No. 70645
Prostaglandin F_{2α} - Item No. 16010
Prostaglandin G₂ - Item No. 17010
Prostaglandin H₂ - Item No. 17020
Prostaglandin Screening EIA Kit - Item No. 514012
UltraPure Water - Item No. 400000

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Said refund or replacement is conditioned on Buyer giving written notice to Cayman within thirty (30) days after arrival of the material at its destination. Failure of Buyer to give said notice within thirty (30) days shall constitute a waiver by Buyer of all claims hereunder with respect to said material.

For further details, please refer to our Warranty and Limitation of Remedy located on our website and in our catalog.

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NOTES

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