Antiserum was developed in rabbits using purified recombinant human CYP1A2 as immunogen. The whole IgG fraction was purified from antiserum using caprylic acid/ammonium sulfate fractionation. Anti-CYP1A2 IgG is provided as a powder after lyophylization from 100 mM potassium phosphate buffer (pH 7.4), 150 mM KCl, and 2.5 µM thimerosal (added as a preservative).

**Specificity and Purity**
Specificity has been determined by Western blotting. Anti-CYP1A2 IgG reacts with both CYP1A2 and CYP1A1 (see below), the former of which is found in human liver while the latter is expressed in extrahepatic tissues only.

Antibody purity has been established by SDS-PAGE run under denaturing conditions. Two protein bands with molecular weights of 50 kDa and 25 kDa can be visualized by Coomassie blue staining, which correspond to the heavy and light chains, respectively, of rabbit IgG.

**Reconstitution of Lyophylized Product and Storage**
Store the lyophylized product at 0-5°C. For Western blotting, reconstitute by adding 1 ml of PBS/50% glycerol to one vial of lyophilized IgG (1 mg) and mix vial gently until powder dissolves. Upon reconstitution, solution can be stored at -20°C, as the presence of glycerol will prevent freeze/thaw cycles. Anti-CYP1A2 IgG solutions without glycerol should be also be stored at -20°C but subjected to freeze/thaw cycles as seldom as possible.

**Use for Western Blotting**
Incubate blots overnight with 2.5 - 5.0 µg rabbit anti-human CYP1A2 IgG/ml of appropriate blocking solution. After washing to remove unbound CYP1A2 antibody, incubate with an anti-rabbit IgG conjugate of choice (e.g., anti-rabbit IgG-peroxidase or anti-rabbit IgG-biotin), and develop accordingly. A detailed Western blotting method can be found in the PROTOCOLS section.

**Use for Immunoinhibition**
Incubation of anti-human CYP1A2 IgG with human liver microsomes at a ratio of 5 mg IgG/nmol microsomal P450 (ca 1.7 mg IgG/mg microsomal protein) before reaction initiation will typically give 80% inhibition of an exemplary CYP1A2-catalyzed reaction (e.g., phenacetin O-deethylation; see attached). The methodology for conducting P450 immunoinhibition assays is given in the PROTOCOLS section.
Antibodies to human CYP1A2 had a marked inhibitory effect on phenacetin O-deethylation by liver microsomes from subject UC9405 (80% inhibition at 5 mg IgG/nmol P450) whereas the other P450 antibodies tested failed to decrease this CYP1A2-catalyzed reaction (Panel A). Maximal inhibition (80%) of microsomal phenacetin oxidation in this human subject was achieved at an anti-CYP1A2 IgG:P450 ratio of 7.5 mg/nmol (Panel B).